

HOSTED BY



Contents lists available at ScienceDirect

Journal of Traditional and Complementary Medicine

journal homepage: <http://www.elsevier.com/locate/jtcm>

Original article

Efficacy of Dragon's blood cream on wound healing: A randomized, double-blind, placebo-controlled clinical trial



Foroogh Namjoyan ^{a,*}, Fatemeh Kiashi ^b, Zahra Beigom Moosavi ^c, Fatemeh Saffari ^b, Behzad Sharif Makhmalzadeh ^d

^a Pharmacognosy Department, Marine Natural Pharmaceutical Research Center, School of Pharmacy, Jundishapur University of Medical Sciences, Ahvaz, Iran

^b School of Pharmacy, Jundishapur University of Medical Sciences, Ahvaz, Iran

^c Dermatology Department, School of Medicine, Jundishapur University of Medical Sciences, Ahvaz, Iran

^d Pharmaceutics Department, School of Pharmacy, Jundishapur University of Medical Sciences, Ahvaz, Iran

ARTICLE INFO

Article history:

Received 16 July 2014

Received in revised form

27 August 2014

Accepted 16 September 2014

Available online 22 January 2015

Keywords:

clinical trial

Dragon's blood

wound healing

ABSTRACT

The blood-red sap of Dragon's blood has been used in folk medicine for fractures, wounds, inflammation, gastrointestinal disorders, rheumatism, blood circulation dysfunctions, and cancer. Existing *in vitro* and *in vivo* bioactivity of this herb on different mechanisms of healing shows strong potential of this sap in wound healing. This clinical trial study was designated to evaluate the wound healing effect of Dragon's blood on human wounds. Sixty patients, between the ages of 14–65 years, who were referred to remove their skin tag, were assigned to this double-blind, placebo-controlled, randomized clinical trial and received either Dragon's blood or a placebo cream. They were visited on the 3rd, 5th, 7th, 10th, 14th, and 20th day of the trial to check the process of healing and to measure the wound's surface. At the end of trial, there was a significant difference in the mean duration of wound healing between the two groups ($p = 0.0001$). The phenolic compounds and the alkaloid taspine, which exist in Dragon's-blood resin, are probably the main reasons for the wound healing property of this plant. Being natural accessible, safe, and affordable makes Dragon's blood cream, a good choice for addition to the wound healing armamentarium. Further studies on wounds with different causes and among larger populations are suggested to ensure the effectiveness and safety of Dragon's blood.

Copyright © 2014, Center for Food and Biomolecules, National Taiwan University. Production and hosting by Elsevier Taiwan LLC. All rights reserved.

1. Introduction

Dragon's blood, a deep red resin, is a well-known traditional medicine, obtained from four different sources: *Croton* spp. (syn. Sangre de draco, Euphorbiaceae), *Dracaena* spp. (syn. Zanzibar drop, Dracaenaceae), *Daemonorops* spp. (syn. Jerang or Djerang, Palmaceae), and *Pterocarpus* spp. (syn. East India Kino or Malabar Kino, Fabaceae). Dragon's blood has been used by different civilizations such as the Greeks, the Romans, and the Arabs.¹ *Croton lechleri* has several medicinal properties, such as wound healing,^{1,2} cicatrizing,³ immunomodulator,^{1,4} analgesic, antiulcer,

antidiarrheal,¹ antibacterial,⁵ antiviral,⁶ antihemorrhagic,¹ anti-inflammatory, antioxidant,^{1,3} mutagenic and antimutagenic,^{1,3,7} antitumor,^{1,8} anticancer,⁹ and cytotoxic effects.^{1,3} Proanthocyanidins are the main chemical constituent of the resin, >90% of the dry weight.¹⁰ It also contains taspine, an alkaloid, and catechin, epigallocatechin, epicatechin, and a small percentage of terpene compounds.^{11,12}

Wounds, the physical damage to the skin and its underlying structure, can result from trauma, burns, or chemicals.¹³ Wound healing is a complicated process. According to cellular and molecular mechanisms, there are three overlapping phases of wound healing as follows: inflammation due to the migration of fibroblasts and inflammatory cells, such as neutrophils and monocytes, into the wound site; then, reconstruction of the epithelial barrier and production matrix at the site of injury leading to new tissue formation; and finally, maturation.^{14–16}

* Corresponding author. Pharmacognosy Department, School of Pharmacy, Ahvaz Jundishapur University of Medical Sciences, Golestan Ave., Ahvaz, Iran.

E-mail addresses: namjoyan_F@ajums.ac.ir, namjoyan@yahoo.com (F. Namjoyan).

Peer review under responsibility of The Center for Food and Biomolecules, National Taiwan University.

Dragon's blood has the immunomodulatory property by influence on complement system.⁴ The antioxidant agents from Dragon's blood leaves and fruit, such as phenolic profile and organic acids, are able to protect against free radicals.^{17,18} Noteworthy, the anti-inflammatory effect of the alkaloid taspine has been reported.¹⁹ These studies have shown this plant can promote healing by affecting the inflammatory phase. Studies confirm that after only 1 day of treatment with Dragon's blood, the wound contracts and a dark crust forms on the wound surface which prevents secondary infection.^{2,20} It also stimulates the proliferation and migration of fibroblasts and the production of collagen, resulting in epithelial regeneration and wound healing which can affect second and third phases of the healing process.^{2,12}

Although there are some studies addressing the healing effect of Dragon's blood on cell lines in animal models,^{2,20,27} to best of our knowledge, the effect on human skin has not been studied yet. In this study, we designated to evaluate the healing effect of Dragon's blood on human skin.

2. Materials and methods

2.1. Trial design

This double-blind, placebo-controlled, randomized clinical trial was conducted on 60 patients referred to the dermatology clinic of Imam Khomeini Hospital (Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran.) for the removal of their skin tags between October 2010 to November 2011. Written consent was obtained from all participants. The trial was registered by the IRCT Iranian Registry of Clinical Trials (IRCT201008224610N1), in accordance with the Helsinki Declaration of 1975, and was confirmed by the Ethics Committee of Ahvaz Jundishapur University of Medical Sciences (Eth NO. 715).

The patients, aged between 14 years and 65 years, were referred for skin tag removal using cautery. Inclusion criteria were lesions between 3 mm and 10 mm in diameter and exclusion criteria were uncontrolled or chronic diseases, pregnancy, or breastfeeding. Demographic characteristics such as sex, age, and skin type, and number and anatomical area of lesions were recorded in the questionnaire. After measuring the wound's surface using checkered transparent paper (the surface of each square was 1 mm²), a therapeutic or placebo cream was given to the patients randomly (according to block method randomization). All patients were asked to use the cream twice a day and store it at room temperature. They were also asked not to use any other medication for wound healing. The wound's surface and recovery was considered 100% and 0%, respectively, on the 1st day. The patients were visited on the 3rd, 5th, 7th, 10th, 14th, and 20th day of the trial to check the healing process and to measure the wound's surface. If recovery was not 100% on the 20th day, they were followed up until the wound's surface and recovery were 0% and 100%, respectively.

2.2. Plant material

Croton lechleri powder was purchased from the Maya Ethnobotanicals, Harleem, Netherland Company. Extraction was performed using a Soxhlet apparatus with ethanol (80%) for 4 hours,¹² and was then filtrated and concentrated using a rotary evaporator (Heidolph, Germany). Finally, the extract was dried in a freeze drier (Operon, Korea).

2.3. Phytochemical study

Total phenolic content was determined using the Folin-Ciocalteu method.²¹ Briefly, to 0.5 mL of each sample (tannic acid

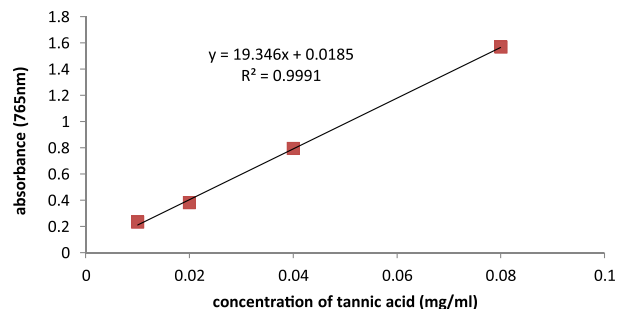


Fig. 1. Tannic acid Standard curve in measurement of total phenolic content using Folin- Ciocalteu method.

as positive standard or extract) 2.5 mL 1/10 diluted Folin–Ciocalteu reagent was added. After 5 minutes, 2 mL of Na₂CO₃ (7.5% w/v) was added and incubated at room temperature in a dark place for 2 hours. The absorbance of all samples was measured at 765 nm. The results were expressed as g of tannic acid equivalent/100 g of dry extract powder.

2.4. Preparation of Dragon's blood cream

The formulation was as follows: 10% cetyl alcohol, 7% isopropylmeristat, and 21% Vaseline(cream base); 1.5% span20 and 1.5% tween80 (emulsifying agent); 0.02% propylparaben and 0.18% methylparaben (preservative); 5% propylene glycol (humectant); 15% ethanolic extract of *C. lechleri*, and distilled water.

The same ingredients were used to prepare the placebo cream with the exception of the herbal extract. Permitted food colors were used to achieve the color of the therapeutic cream (reddish brown). Similar tubes were filled with the therapeutic and placebo cream and labeled. Both creams were differentiated using codes that were unknown to the participants and researcher.

2.5. Statistical analysis

The data were expressed as mean ± standard deviation and analyzed using the two-tailed Student *t* test to compare the mean duration of wound healing between the two groups and the percentage of wound healing per day, and the analysis of variance (ANOVA) method was used to compare the mean duration of healing between different anatomical areas. A value of *p* < 0.05 was considered significant.

3. Results

The presence of alkaloid was confirmed by black spot in Dragandrof, and cream and brown precipitation in Mayer and Wagner reagents, respectively.

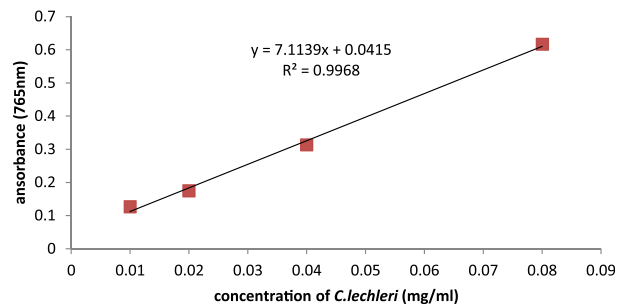


Fig. 2. Measurement of total phenolic content of *C. lechleri* using Folin- Ciocalteu method.

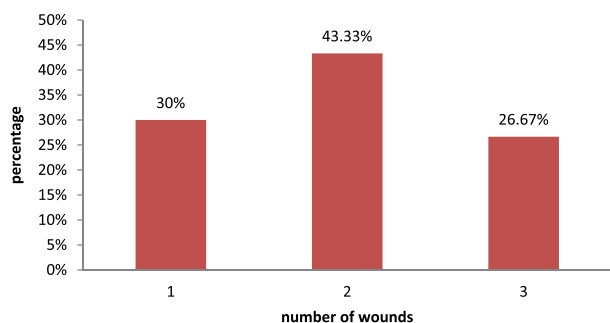


Fig. 3. Number of wounds in studied population.

Total phenolic content was measured using the Folin–Ciocalteu method and tannic acid was used as a positive standard (Figs. 1 and 2).

Tannic acid equivalent in 100 g Dragon's blood powder was 6.632 ± 0.65 g, and in 100 g dry extract was 43.35 ± 4.27 g. The prepared cream contained 15% extract, therefore, it included 6.5025 g total phenolic compounds.

Sixty participants (31 women and 29 men) entered into the trial. Eighteen patients had one wound, 26 patients had two wounds, and 16 patients had three wounds (Fig. 3). We investigated 100 wounds in total. Forty-five wounds were assigned to the therapeutic group and 55 wounds were assigned in the placebo group randomly. Thirty-eight wounds (63.34%) were on the neck, 20 wounds (43.33%) were on the trunk, and 16 wounds (26.67%) were on the lower limbs. There were no significant differences between the two groups for age ($p = 0.799$) and wound surface ($p = 0.946$) before starting the study (Tables 1 and 2). Twelve patients (17 wounds) had poor compliance and withdrew from the study. There were no irritations or wound infections among the therapeutic participants, whereas one patient in the placebo group showed wound infection, and was treated with mupirocin topical ointment and dropped out of the study. Eventually, the trial was completed with 37 wounds in the therapeutic group and 46 wounds in the placebo group.

Two-tailed Student *t* test was used to compare the mean duration of healing and the percentage of wound healing between the two groups. The differences in the percentage of wound healing between the two groups were significant on all days of the trial (Table 3).

To compare the mean duration of healing between different anatomical areas, the ANOVA test was used. There were no significant differences between anatomical areas in both groups ($p > 0.05$).

Table 1
Comparison of mean age between two groups before starting the study.

Group	Mean of age	SD	P value
Therapeutic	38.4	10.98	0.799
Placebo	48.96	10.99	0.799

Table 2
Comparison of wound's surface between two groups before starting the study.

Group	Number of wounds	Mean of wound's surface	SD	P value
Therapeutic	45	8.54	4.1	0.946
placebo	55	8.6	4.048	0.946

Table 3
The mean percentage of wound healing in different days for two groups.

	3 rd day	5 th day	7 th day	10 th day	14 th day
Therapeutic	31.06%	63.77%	77.80%	89.14%	95.73%
Placebo	4.74%	23.50%	43.90%	61.95%	78.10%
P value	0.0001	0.0001	0.0001	0.0001	0.004

4. Discussion

The results of this clinical trial showed that Dragon's blood cream can significantly improve the wound healing duration ($p = 0.0001$).

Previous studies reported the antioxidant and anti-inflammatory effects of Dragon's blood sap.^{1,3,22} It is also reported to inhibit the lipid peroxidation in the liver of mice.²³ In this study, we observed a significant improvement of wound healing from the 3rd day, which may be due to a shortening of the inflammation process because of the presence of phenolic compounds such as proanthocyanidins and catechin.^{1,20}

Our results are in agreement with two studies which investigated the healing effect of Dragon's blood on rats. One of these studies showed the wound healing effect of the alkaloid taspine in rats. It suggested taspine stimulated chemotaxis of fibroblasts.²⁴ Another study showed the wound healing activity of Dragon's blood was because of the high percentage of polyphenolic compounds in this plant.^{1,2}

Inflammation is usually a suitable media for infections, and healing is delayed in these situations.²⁵ The polyphenolic compounds of the sap create a protective layer on the wound surface, and this physical barrier prevents microbial contamination. Some compounds of the resin, 2,4,6-trimethoxyphenol, 1,3,5-trimethoxybenzene, crolechinic acid, and korberin, which showed antibacterial effects, can indirectly improve the healing process.^{1,20}

Wound contraction is one of the factors that facilitates re-epithelialization.²⁵ The polyphenolic compounds condense and clog the wounds by binding to proteins, and enzymes lead to ailment.²⁰ Frolidi et al²⁶ have suggested that the vasoconstriction effect of Dragon's blood affects wound healing probably by fastening the wound.

Angiogenesis and matrix reformation phases occur after re-epithelialization. The migration and proliferation of fibroblasts causes reformation of the matrix.²⁵ Dragon's blood sap probably affects wound healing through increasing the migration of human fibroblasts in the cell culture and proliferation of the endothelial cells.^{12,24}

5. Conclusion

This clinical trial suggests Dragon's blood is a potent, available, affordable, and safe healing agent. The exact role of Dragon's blood in the pathogenesis of wound healing regarding its effect on stimulation or hindering mediator's synthesis is still absent and further studies are required. Our study encourages evaluating the healing process on other wounds such as diabetic ulcers, bedsores, or burns.

References

- Gupta D, Bleakley B, Gupta RK. Dragon's blood: botany, chemistry and therapeutic uses. *J Ethnopharmacol.* 2008;115:361–380.
- Pieters L, De Bruyne T, Van Poel B, et al. *In vivo* wound healing activity of Dragon's blood (*Croton spp.*), a traditional South American drug, and its constituents. *Phytomedicine.* 1995;2:17–22.
- Lopes MI, Saffi J, Echeverrigaray S, Henriques JAP, Salvador M. Mutagenic and antioxidant activities of *Croton lechleri* sap in biological systems. *J Ethnopharmacol.* 2004;95:437–445.

4. Risco E, Ghia F, Vila R, Iglesias J, Álvarez E, Cañigueral S. Immunomodulatory activity and chemical characterisation of sangre de drago (dragon's blood) from *Croton lechleri*. *Planta Med.* 2003;69:785–794.
5. Edvard HGM, de Oliveira LFC, Quye A. Raman spectroscopy of coloured resins used in antiquity: dragon's blood and related substances. *Spectrochim Acta A Mol Biomol Spectrosc.* 2001;57:2831–2842.
6. Ubillas R, Jolad SD, Bruening RC, et al. SP-303, an antiviral oligomeric proanthocyanidin from the latex of *Croton lechleri* (sangre de drago). *Phytomedicine.* 1994;1:77–106.
7. Rossi D, Guerrini A, Paganetto G, et al. *Croton lechleri* Müll Arg. (Euphorbiaceae) stem bark essential oil as possible mutagen-protective food ingredient against heterocyclic amines from cooked food. *Food Chem.* 2013;139:439–447.
8. Alonso-Castro AJ, Ortiz-Sánchez E, Domínguez F, et al. Antitumor effect of *Croton lechleri* Mull. Arg. (Euphorbiaceae). *J Ethnopharmacol.* 2012;140:438–442.
9. Montopoli M, Bertin R, Chen Z, Bolcato J, Caparrotta L, Frolidi G. *Croton lechleri* sap and isolated alkaloid taspine exhibit inhibition against human melanoma SK23 and colon cancer HT29 cell lines. *J Ethnopharmacol.* 2012;144:747–753.
10. Cai Y, Evans FJ, Roberts MF, Phillipson JD, Zenk MH, Gleba YY. Polyphenolic compounds from *Croton lechleri*. *Phytochemistry.* 1991;30:2033–2040.
11. Phillipson JD. A matter of some sensitivity. *Phytochemistry.* 1995;38:1319–1343.
12. Vaisberg AJ, Milla M, Planas MC, Cordova JL, de Agusti ER, Ferreyra R, Mustiga MC, Carlin L, Hammond GB. Taspine is the Cicatrizant Principle in Sangre de Grado Extracted from *Croton lechleri*. *Planta Med.* 1989;55:140–143.
13. Martindale. *The complete drug reference.* 36th ed. United Kingdom: Pharmaceutical Press; 2009.
14. Jia Y, Zhao G, Jia J. Preliminary evaluation: the effects of *Miller* and *Aloe arborescens* Miller on wound healing. *J Ethnopharmacol.* 2008;120:181–189.
15. Ashcroft GS, Roberts AB. Loss of Smad3 modulates wound healing. *Cytokine Growth Factor Rev.* 2000;11:125–131.
16. Wang AS, Armstrong EJ, Armstrong AW. Corticosteroids and wound healing: clinical considerations in the perioperative period. *Am JSurg.* 2013;206:410–417.
17. Santos RP, Mendes LS, Silva BM, et al. Phytochemical profiles and inhibitory effect on free radical-induced human erythrocyte damage of *Dracaena draco* leaf: a potential novel antioxidant agent. *Food Chem.* 2011;124:927–934.
18. Silva BM, Santos RP, Mendes LS, et al. *Dracaena draco* L. fruit: phytochemical and antioxidant activity assessment. *Food Res Int.* 2011;44:2182–2189.
19. Perdue GP, Blomster RN, Blake DA, Farnsworth NR. South American plants II: taspine isolation and anti-inflammatory activity. *J Pharm Sci.* 1979;68:124–125.
20. Chen ZPCY, Phillipson JD. Studies on the anti-tumor, anti-bacterial, and wound-healing properties of Dragons blood. *Planta Medica.* 1994;60:541–545.
21. Singleton VL, Orthofer R, Lamuela-Raventós RM. [14] Analysis of total phenols and other oxidation substrates and antioxidants by means of folin-ciocalteu reagent. In: Lester P, ed. *Methods in Enzymology.* vol. 299. Academic Press; 1999:152–178.
22. Desmarchelier C, Schaus FW, Coussio J, Cicca G. Effects of Sangre de Drago from *Croton lechleri* Muell.-Arg. on the production of active oxygen radicals. *J Ethnopharmacol.* 1997;58:103–108.
23. Desmarchelier CJ, de Moraes Barros SB. Pharmacological activity of South American plants: effects on spontaneous in vivo lipid peroxidation. *Phytother Res.* 2003;17:80–82.
24. Porras-Reyes BH, Lewis WH, Roman J, Simchowicz L, Mustoe TA, eds. *Enhancement of wound healing by the alkaloid taspine defining mechanism of action. Proceedings of the Society for Experimental Biology and Medicine Society for Experimental Biology and Medicine.* New York, NY: Royal Society of Medicine; 1993.
25. McGibbon D. Rook's Textbook of Dermatology. *Clin Exp Dermatol.* 2006;31:178–179.
26. Frolidi G, Zagotto G, Filippini R, Montopoli M, Dorigo P, Caparrotta L. Activity of sap from *Croton lechleri* on rat vascular and gastric smooth muscles. *Phyto-medicine.* 2009;16:768–775.
27. He Xuan-ling, Wang Shen-zhi, Huang Zheng-de. Effects of dracaena on expression of Smads protein in skin ulcers in diabetic rats. *J Trad Chin Med Univ Hunan.* 2010–11.